# Functional Characterization of All Missense Variants in LEPR, PCSK1, and POMC Genes Arising From Single-Nucleotide Variants

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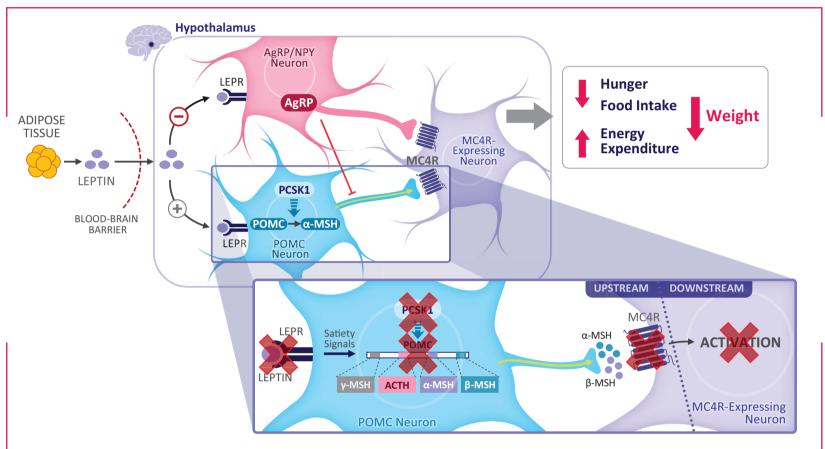
## Summary

- variants [SNVs]) in leptin receptor (*LEPR*), proprotein convertase subtilisin/kexin type 1 (*PCSK1*), and proopiomelanocortin (*POMC*)

# Introduction

- Rare genetic disorders of obesity are characterized by early-onset, severe obesity and hyperphagia
- Rare genetic disorders of obesity can be caused by LOF variants in genes composing the melanocortin 4 receptor (MC4R) pathway, including LEPR, PCSK1, and POMC<sup>1</sup> (Figure 1)

Figure 1. The melanocortin 4 receptor pathway, a component of the central melanocortin pathway, helps regulate appetite, body weight, and energy expenditure, and loss-of-function variants in this pathway can cause rare genetic disorders of obesity.<sup>1</sup>



ACTH, adrenocorticotropic hormone: AgRP, agouti-related protein: LEPR, leptin receptor: MC4R, melanocortin 4 receptor: MSH, melanocytestimulating hormone; NPY, neuropeptide Y; PČSK1, proprotein convertase subtilisin/kexin type 1; POMC, proopiomelanocortin.

• To improve the understanding of the genetics underlying rare genetic disorders of obesity, it is crucial to characterize the impact of observed genetic variants on protein function

# **Objective**

To perform biochemical characterization and determine functionality of all conceivable missense variants arising from SNVs in LEPR, PCSK1, and POMC

# Methods

### LEPR Assay<sup>5</sup>

- The functionality of LEPR variants was assessed using a LEPR-STAT3-luciferase assay, given that LEPR-STAT3 signaling is critical for leptin-regulated appetite and body weight (Figure 2A)
- Human embryonic kidney (HEK) cells were transiently transfected with relevant LEPR variants, firefly luciferase (LEPR-STAT3 responsive), and Renilla luciferase (transfection control)
- 48 hours after transfection, cells were serum starved for 6 hours and subsequently treated with leptin (10 ng/mL) • 6 hours after leptin treatment, the media were removed and the cell lysate was prepared using Dual-Glo<sup>®</sup> lysis buffer (Promega Corporation, Madison, WI)
- The cell lysate was transferred to a 384-well plate and treated with Dual-Glo luciferase reagents according to the manufacturer's instructions
- Renilla and firefly luminescence were measured and the luminescence ratio was calculated on an EnVision plate reader (PerkinElmer Inc, Waltham, MA)

### PCSK1 Assay<sup>6</sup>

- The functionality of *PCSK1* variants was investigated by measuring the processing of a fluorogenic substrate containing a PCSK1 cleavage site (Figure 2B)
- HEK293T cells were transiently transfected with PCSK1 (wild-type [WT] or missense variants)
- To determine the PCSK1 catalytic activity, conditioned media from transfected cells were subjected to a fluorogenic assay

- were taken over 2 hours on an EnVision plate reader LEPR and SRE Add leptin luciferase (10 ng/mL) to transfection in transfected cells HEK cells PCSK Conditioned media from transfected transfection in HEK cells cells (PCSK1) РОМС transfection transfected in HEK cells cells (α- <u>&</u> β-MSH α- <u>or</u> β-MSH expressing peptides **POMC** Assay (cAMP) assay (Figure 2C) POMC variants were transfected in HEK cells
- After 1 hour, cAMP expression was guantified
- (Figure 2D)
- After 1 hour, cAMP expression was quantified

### Variant Classification

- validated against the published functionality of each variant
- categorized

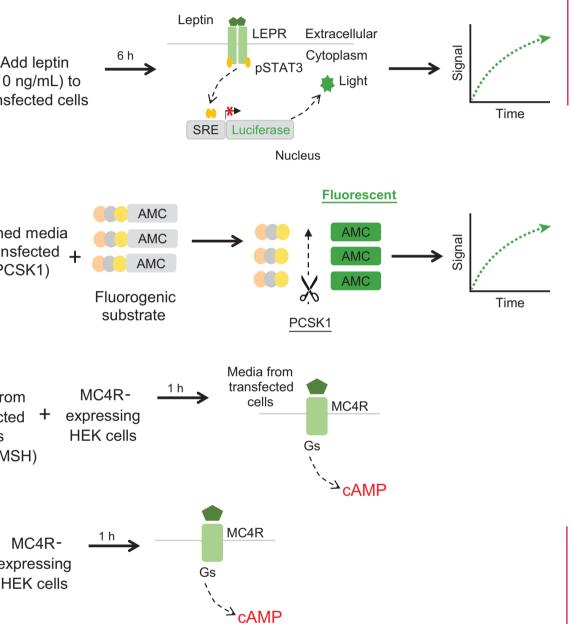
# • To determine the functional effects of genetic variants on protein function and thus their potential contribution to MC4R pathway-related obesity, we performed functional characterization of all conceivable missense variants (arising from single-nucleotide)

• Of all conceivable missense variants arising from SNV, 18.7% of LEPR variants, 45.4% of PCSK1 variants, and 8.3% of POMC variants exhibited varying degrees of loss of function (LOF)

PCSK1 activity was evaluated in triplicate in 50-µL reactions in a 96-well plate containing 25 µL of conditioned media and 25 µL of buffer containing final concentrations of 200 µM of substrate pyr-ERTKR-AMC, 100 mM of sodium acetate (pH 5.5), 2 mM of CaCl<sub>2</sub>, 0.1% Brij 35, and 1× of protease inhibitor cocktail

Reaction mixtures were incubated at 37°C and fluorescence measurements (380 nm excitation, 460 emission)

Figure 2. Biochemical assays for (A) LEPR, (B) PCSK1, and (C, D) POMC variants



AMC, 7-amino-4-methylcoumarin; cAMP, cyclic adenosine monophosphate; HEK, human embryonic kidney; LEPR, leptin receptor; MC4R, melanocortin 4 receptor; MSH, melanocyte-stimulating hormone; PCSK1, proprotein convertase subtilisin/kexin type 1; POMC, propriomelanocortin; pSTAT3, phosphorylated signal transducer and activator of transcription 3; SRE, serum response element.

• The functional effect of *POMC* variants was characterized using an MC4R–cyclic adenosine monophosphate

• A transgene-based approach assessed the functional effect of *POMC* variants outside the  $\alpha/\beta$ –MSH region

• Media (α- and β-MSH) from transfected cells were collected 48 hours later and combined with MC4R-expressing HEK cells

• A peptide-based approach assessed the functional effect of *POMC* variants inside the  $\alpha/\beta$ -MSH region

•  $\alpha$ - or  $\beta$ -MSH peptides for each missense variant (arising from SNV) were synthesized

• Each α- or β-MSH peptide was combined with MC4R-expressing HEK cells

Variants were classified as significant LOF, moderate LOF, or WT-like on the basis of specific criteria (Table 1) Published variants in *LEPR*, *PCSK1*, and *POMC* underwent biochemical assays, the results of which were

• Every conceivable single missense variant in *LEPR*, *PCSK1*, and *POMC* was analyzed and functionally

• A subset of variants included those identified from genomic databases and previous publications

### Table 1. Criteria for Variant Classification

Gene	Significant LOF	Moderate LOF	WT-like			
LEPR	≤30% of WT	31%-70% of WT	≥71% of WT			
PCSK1	≤30% of WT	31%-70% of WT	≥71% of WT			
POMC (outside $\alpha/\beta$ –MSH)	E <sub>max</sub> ≤30% of WT	E <sub>max</sub> 31%-60% of WT	E <sub>max</sub> ≥61% of WT			
POMC (inside $\alpha/\beta$ –MSH)	$E_{max} \leq 30\%$ of WT <u>or</u> EC <sub>50</sub> >20x (WT EC <sub>50</sub> )	E <sub>max</sub> 31%-60% of WT <u>or</u> EC <sub>50</sub> (WT EC <sub>50</sub> + 3SD) – (20 x [WT EC <sub>50</sub> ])	$E_{max} \ge 61\%$ of WT and EC <sub>50</sub> < (WT EC <sub>50</sub> + 3SD)			
EC <sub>50</sub> , half-maximal effective concentration; E <sub>max</sub> , maximum effect; LEPR, leptin receptor; LOF, loss of function; MSH, melanocyte-stimulating hormon PCSK1, proprotein convertase subtilisin/kexin type 1; POMC, proopiomelanocortin; SD, standard deviation; WT, wild-type.						
Results						

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### Validation of Biochemical Assays Using Published Genetic Variants

Results of each assay were largely consistent with previously published genetic variants (Table 2) in LEPR,<sup>8-12</sup> PCSK1,<sup>13-19</sup> and POMC<sup>20-2</sup>

Table 2. Validation of Assays Against Published Variants of Known Functional Impact

Gene	Variant	Published data <sup>a</sup>	Rhythm data (% of WT activity)	Validation
LEPR	GFP_Control	NA	1.84	
	W664R <sup>®</sup>	Significant LOF	Moderate LOF (53.21)	No
	A409E <sup>8</sup>	Significant LOF	Significant LOF (-3.78)	Yes
	H684P <sup>8</sup>	Significant LOF	Significant LOF (13.58)	Yes
	R612H <sup>8</sup>	Moderate LOF	WT-like (81.28)	No
	P878S <sup>®</sup>	Significant LOF	Significant LOF (-2.56)	Yes
	P876S <sup>°</sup>	Significant LOF	Significant LOF (-2.40)	Yes
	Q270P <sup>10</sup>	Significant LOF	Significant LOF (6.75)	Yes
	Y1141F <sup>11</sup>	Significant LOF	Significant LOF (-0.64)	Yes
	Q223R <sup>12</sup>	WT-like	WT-like (89.19)	Yes
PCSK1°	GFP_Control	NA	1.29	_
	G226R <sup>13</sup>	Significant LOF	Significant LOF (0.91)	Yes
	G209R <sup>14</sup>	Significant LOF	Significant LOF (1.84)	Yes
	N423K <sup>14</sup>	Significant LOF	Significant LOF (1.28)	Yes
	F548S <sup>14</sup>	Significant LOF	Significant LOF (1.57)	Yes
	G593R <sup>14</sup>	Significant LOF	Significant LOF (1.19)	Yes
	S307L <sup>15</sup>	Significant LOF	Significant LOF (2.09)	Yes
	N309K <sup>16</sup>	Significant LOF	Significant LOF (0.89)	Yes
	T175M <sup>13</sup>	Significant LOF	Significant LOF (6.66)	Yes
	M125I <sup>13</sup>	Significant LOF	Significant LOF (17.73)	Yes
	S24C <sup>17</sup>	WT-like	WT-like (78.34)	Yes
	S690T <sup>18</sup>	WT-like	WT-like (93.84)	Yes
	K26E <sup>13</sup>	WT-like	WT-like (84.97)	Yes
	H72L <sup>19</sup>	WT-like	WT-like (118.21)	Yes
	T558A <sup>13</sup>	WT-like	Moderate LOF (42.89)	No
POMC	GFP_Control	NA	1.30	
	E105 <sup>23,d</sup>	Significant LOF	0	Yes
	Y221C <sup>20,e</sup>	Moderate LOF	Moderate LOF (98.0, 173 nM)	Yes
	F144L <sup>21</sup>	Significant LOF	Significant LOF (23.56, >1 µM)	Yes
	D53G <sup>24</sup>	WT-like	WT-like (95.86)	Yes
	R236G <sup>22,e</sup>	Moderate LOF	WT-like (88.56)	No
	H143Q <sup>20</sup>	Significant LOF	Significant LOF (44.79, >1 µM)	Yes

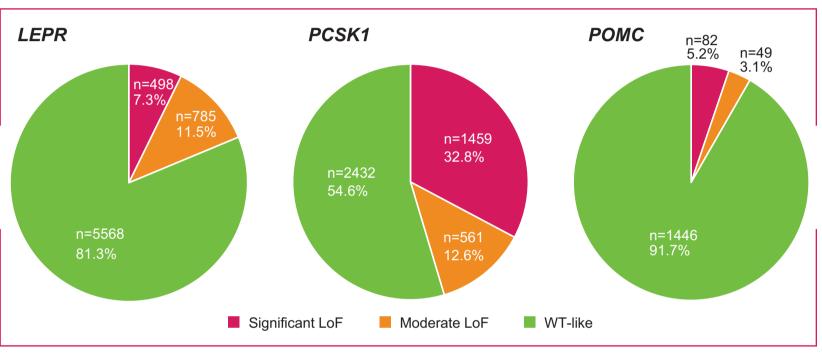
kexin type 1; POMC, proopiomelanocortin; WT, wild-type. "Determined gualitatively on the basis of data in publication." Determined by comparing percentage of WT activity category with published LOF category. Wild-type activity measured at 1 hour. Stop codon (nonsense mutation). Mutation in β-MSH

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### Functional Characterization of Missense Variants Arising From SNV in *LEPR*, *PCSK1*, and *POMC*

- Out of 6851 *LEPR* variants arising from SNV, 498 (7.3%) exhibited significant LOF, 785 (11.5%) exhibited moderate LOF, and 5568 (81.3%) exhibited WT-like activity (Figure 3A)
- Out of 4452 PCSK1 variants arising from SNV, 1459 (32.8%) exhibited significant LOF, 561 (12.6%) exhibited moderate LOF, and 2432 (54.6%) exhibited WT-like activity (Figure 3B)
- Out of 1577 *POMC* variants arising from SNV, 82 (5.2%) exhibited significant LOF, 49 (3.1%) exhibited moderate LOF, and 1446 (91.7%) exhibited WT-like activity (Figure 3C)

Figure 3. Functional characterization of all missense variants arising from SNV in LEPR, PCSK1, and POMC.



LEPR, leptin receptor; LOF, loss of function; PCSK1, proprotein convertase subtilisin/kexin type 1; POMC, proopiomelanocortin; SNV, single-nucleotide variant: WT. wild-type

### Functional Characterization of Observed Missense Variants in *LEPR*, *PCSK1*, and *POMC*

- Out of 672 observed LEPR missense variants, 38 (5.7%) exhibited significant LOF, 98 (14.6%) exhibited moderate LOF, and 536 (79.8%) exhibited WT-like activity (Table 3)
- Out of 449 observed PCSK1 missense variants, 138 (30.7%) exhibited significant LOF, 100 (22.3%) exhibited moderate LOF, and 211 (47.0%) exhibited WT-like activity
- Out of 233 observed POMC missense variants, 15 (6.4%) exhibited significant LOF, 9 (3.9%) exhibited moderate LOF, and 209 (89.7%) exhibited WT-like activity

### Table 3. Functional Characterization of Observed<sup>®</sup> Missense Variants in LEPR, PCSK1, and POMC

	<i>LEPR</i> (n=672) n (%)	<i>PCSK1</i> (n=449) n (%)	<i>POMC</i> (n=233) n (%)
Significant LOF	38 (5.7)	138 (30.7)	15 (6.4)
Moderate LOF	98 (14.6)	100 (22.3)	9 (3.9)
WT-like	536 (79.8)	211 (47.0)	209 (89.7)
LEPR, leptin receptor; LOF, los	s of function; PCSK1, proprotein conve	ertase subtilisin/kexin type 1; POMC, pro	opiomelanocortin. <sup>a</sup> As reported in public

databases (gnomAD, 1000 Genomes Project, DiscovEHR, NHLBI Exome Sequencing Project v. 6500) and scientific publications.

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References: 1. da Fonseca et al. J Diabetes Complications. 2017;31:1549-1561. 2. Yazdi et al. PeerJ. 2015;3:e856. 3. Krashes et al. *Nat Neurosci.* 2016;19:206-219. **4.** Cone. *Endocr Rev.* 2006;27:736-749. **5.** Kimber et al. *Endocrinology.* 2008;149:6043-6052. **6.** Blanco et al. Endocrinology. 2015;156:3625-3637. 7. Clement et al. Nat Med. 2018;24:551-555. 8. Farooqi et al. N Engl J Med. 2007;356:237-247. 9. Couturier and Jockers. J Biol Chem. 2003;278:26604-26611. 10. Crouse et al. J Biol Chem. 1998;273:18365-18373. 11. Baumann et al. Proc Natl Acad Sci U S A. 1996;93:8374-8378. 12. Stratigopoulos et al. Obesity (Silver Spring). 2009;17:126-135. 13. Creemers et al. Diabetes. 2012;61:383-390. 14. Martin et al. Gastroenterology. 2013;145:138-148. 15. Farooqi et al. J Clin Endocrinol Metab. 2007;92:3369-3373. **16.** Wilschanski et al. *PLoS One.* 2014;9:e108878. **17.** Loffler et al. *Mol Metab.* 2017;6:295-305. **18.** Mbikay et al. *Mol Genet Metab.* 2011;104:682-687. 19. Williamson et al. J Biol Chem. 2015;290:23214-23225. 20. Lee et al. Cell Metab. 2006;3:135-140. 21. Dubern et al. Pediatr Res. 2008;63:211-216. 22. Challis et al. Hum Mol Genet. 2002;11:1997-2004. 23. Koks et al. Int J Neuropsychopharmacol. 2006;9:167-174. **24.** Cantara et al. AACE Clin Case Rep. 2019;5:e132-e137.